

Microstructural changes and the effect on myofibril proteins in yamu (*Brycon amazonicus*) fish meat during cold storage

Cambios microestructurales y efecto sobre las proteínas miofibrilares de la carne del pescado yamú (*Brycon amazonicus*) durante la conservación en frío

Daniel Castañeda¹, José Manuel Lozano², and Héctor Suárez³

ABSTRACT

This study aimed to evaluate the effect of cold storage on yamu fish meat (*Brycon amazonicus*). The methodology consisted of analyzing two temperatures ($0\pm 1.6^{\circ}\text{C}$ and $-8\pm 1^{\circ}\text{C}$), two storage periods (13 and 61 hours) and two different sources (farm and river), and determine changes in the protein profile, microstructural, physical and chemical properties (water retention capacity-WRC, pH, texture of yamu meat filets). The microstructural changes were observed with optical and scanning electron microscopy. The results of the image analysis of the meat microstructure showed a negative effect from cold storage on the muscle, affecting the connective tissue. Otherwise, the analysis of protein profile demonstrated protein degradation during the cold storage of the yamu meat, such as for heavy chain myosin, actin and α -actinin. In addition, the origin of the fish and the storage temperature had a significant effect on the texture changes ($P\leq 0.05$); meanwhile, the storage time had a significant effect on the three technological characteristics of the meat (WRC, pH and texture) ($P\leq 0.05$). The storage temperature did not significantly affect the WRC ($P>0.05$).

Key words: proteolysis, microscopy, cold storage, texture.

RESUMEN

El propósito de este trabajo consistió en evaluar el efecto de la conservación en frío sobre la carne del pescado yamú (*Brycon amazonicus*). La metodología consistió en analizar bajo dos temperaturas ($0\pm 1,6^{\circ}\text{C}$ y $-8\pm 1^{\circ}\text{C}$), dos periodos de conservación (13 y 61 horas) y dos orígenes distintos (cultivo y río), cambios en el perfil proteico, microestructural, propiedades fisicoquímicas (capacidad de retención de agua-CRA, pH, textura de la carne de filetes de yamú). Las alteraciones microestructurales fueron realizadas por medio de microscopía electrónica de barrido y óptica. Los resultados del análisis de las imágenes de la microestructura de la carne mostraron un efecto negativo de la conservación en frío sobre el músculo, evidenciándose una afectación en el tejido conectivo. Por otro lado, el análisis de perfil proteico permitió evidenciar que durante la conservación en frío de la carne de yamú, proteínas como la miosina de cadena pesada, la actina y la α -actinina presentaron degradación. Adicionalmente, el origen de los peces y la temperatura de almacenamiento tuvieron efecto significativo en los cambios de textura ($P\leq 0.05$) por su parte el tiempo de almacenamiento tuvo efecto significativo sobre las tres características tecnológicas de la carne (CRA, pH y Textura) ($P\leq 0.05$); y la temperatura de almacenamiento no afectó de manera significativa la CRA ($P>0.05$).

Palabras clave: proteólisis, microscopía, almacenamiento en frío, textura.

Introduction

Yamu (*Brycon amazonicus*) is a fish species that has the potential to be farmed because of the meat quality and omnivorous habits that promote food efficiency (Useche *et al.*, 1993). However, the fish farming and supply of yamu meat is still developing although there has been significant progress for issues of reproduction, feeding and cannibalism control at the nursery stage (Arias, 2006), but there are only a few studies on the technological properties and processes that affect yamu meat quality.

It is known that texture is important in determining the quality of fish meat (Ando *et al.*, 1999; Suárez-Mahecha *et al.*, 2007) because losses in this property suggest changes in technological, sensory and microbiological properties. Studies have shown that fish meat loses texture after 24 h of storage on ice (Oka *et al.*, 1990; Toyohara and Shimizu, 1988). The softening of fish meat has been studied in marine fish (Ladrat *et al.*, 2003; Morrison *et al.*, 2000; Wang *et al.*, 2009) and freshwater fish species (Ando *et al.*, 1999; Ma and Yamanaka, 1991; Suárez-Mahecha *et al.*, 2007; Wang *et al.*, 2011); however, these studies have focused on

Received for publication: 29 October, 2016. Accepted for publication: 30 November, 2016.

Doi: 10.15446/agron.colomb.v34n3.61316

¹ Facultad de Ingeniería, Universidad de la Costa. Barranquilla (Colombia).

² Departamento de Farmacia, Facultad de Ciencias, Universidad Nacional de Colombia. Bogota (Colombia).

³ Instituto de Ciencia y Tecnología de Alimentos (ICTA), Universidad Nacional de Colombia. Bogota (Colombia). hsuarezm@unal.edu.co



highly commercial species (Suárez-Mahecha *et al.*, 2007). When stored at low temperatures, yamu meat has rapid softening. Currently, it is not known with certainty which factors cause this or through which mechanisms (Ladrat *et al.*, 2003). Authors have suggested that texture losses in fish meat in cold storage is caused by the effect of proteases (Verrez-Bagnis *et al.*, 1999), especially cathepsins, calpains and hydrolytic enzymes such as elastase and collagenase (Chéret *et al.*, 2007).

It has also been reported that, after storing hake fillets and whole hakes at -8°C for 12 months, there was a substantial increase in the pH (Simeonidou *et al.*, 1997). Another functional property that is affected when fish meat is in cold storage ($<0^{\circ}\text{C}$) is the WRC because it depends on the moisture and stability of the muscular structure (Sánchez-Valencia *et al.*, 2014). Altering this property has negative effects on the marketing of meat because weight loss during storage and during cooking results in immediate rejection by consumers.

The aim of this study was to evaluate the effect of cold storage on yamu meat (*Brycon amazonicus*), depending on temperature, shelf life and origin of fish, on the protein profile, microstructure and technological properties such as the WRC, pH and texture.

Materials and methods

Biological material

The yamu (*Brycon amazonicus*) fishes were obtained in the Department of Meta. One group was obtained from an aquaculture farm located in the department of Meta ($4^{\circ}16'17.1''\text{N}$ and $73^{\circ}29'39.0''\text{W}$, municipality of Cumaral) (farmed); the second group was caught in a natural environment ($4^{\circ}32'13.9''\text{N}$ and $71^{\circ}50'47.2''\text{W}$), municipality of Puerto Gaitan, San Miguel district (fished) (Meta river). The fish (6 from the river and 7 from the farm) were killed with a blow to the head and immediately sent to the laboratory in polystyrene coolers with ice packed in plastic bags, avoiding direct contact between the ice and fish. In the laboratory for meat and derivatives processes, the fish were washed with cold water ($5\pm 1.5^{\circ}\text{C}$), eviscerated and filleted; randomly, fish fillets from each source were assigned to 3 groups of 8 fillets each for subsequent analysis. The first group had fresh samples that were analyzed immediately after filleting. The second group had samples stored at $0\pm 1.5^{\circ}\text{C}$. The third group had samples stored at $-8\pm 1^{\circ}\text{C}$. All of the samples were analyzed during storage for 13 and 61 h.

Microscopic analysis

Optical microscopy

In order to analyze changes in the muscle structure and microstructure of the yamu meat in cold storage, a histological analysis was carried out. The samples were taken at the epaxial level (back) of the fillets and then cut into $2 \times 2 \times 2$ mm pieces, observing the muscle fibers transversely. The pieces were placed in 1.5 mL vials and fixed in 3% buffered formalin for 2 d. They were then dehydrated in a graded series of alcohols (50, 60, 70, 80, 90, and 99%), followed by xylene. The samples used in the optical microscopy were embedded in paraffin and cut into microtome ($4\mu\text{m}$) and stained with Masson trichrome (MT). For this process, the samples were washed with distilled water and immersed in Weigert hematoxylin for 5 min, then washed with tap water at 5°C and immersed in ponceau fuchsin for 5 min and immersed in phosphomolybdic acid for 5 min. This solution was changed to an Aniline Blue solution for 5-7 min. Finally, the tissues were analyzed under a microscope (Leica DM750 P, Leica Microsystems, Heerbrugg, Switzerland) and images were taken at 40X in order to highlight the collagen fibers.

Scanning electron microscopy (SEM)

The samples for this procedure were fixed in 3% glutaraldehyde buffer for 72 h, then dried at the critical point using an EK 3150 for 15 min and covered with gold film using a Quorum, Q150R ES. To proceed with the SEM (FEI, Quanta 200-r), the samples were mounted on individual slides. The images were captured at 4000X.

Water retention capacity (WRC) of the yamu meat samples

The WRC of the yamu meat samples was estimated following the methodology proposed by Sánchez-Alonso (2012), for which 3 g of meat were taken and wrapped in two filter papers (Whatman No. 1, 110 mm diameter) and placed in a falcon tube. Subsequently, they were centrifuged for 15 min at 3,000 g. Then, the paper was removed and the samples were weighed. The WRC was expressed as a percentage of water retained by the sample after centrifugation.

Analysis of the yamu meat texture

The texture was evaluated using a Stable Micro System texture analyzer (TA.XT2, Surrey, UK), with a flat end cylinder (10 mm in diameter, P/10 type) that was pressed against the muscle tissue. All of the tests were done at 5°C , keeping the fillets in Petri dishes on ice. Three cubes ($2 \times 2 \times 1.2$ cm) were taken from each fillet, from the top of the

sideline. The response variable was the maximum cutting force (g g^{-1} sample). A compression test was done before the test: 1.00 mm s^{-1} , test speed: 1.10 mm s^{-1} , speed after test: 10.00 mm s^{-1} , distance between the cylinder and sample: 15.0 mm, compression of the sample: 40.0%. The conditions were the same for each sample and seven measurements were taken for each meat portion (Larsson *et al.*, 2014).

pH determination for the yamu meat samples

This parameter was analyzed following the methodology described by Mohan *et al.* (2007), in which a homogenized sample was held with cold deionized water ($0 \pm 0.5^\circ\text{C}$) 1:6, using a homogenizer (Ultra-Turrax IKA T-25, Campinas, Brazil) at 10,000 rpm for 1 min. Once the homogenate was obtained, the reading was taken with a pH meter ($17 \pm 1.5^\circ\text{C}$) (JENWAY®, JW-3505, Staffordshire, UK). Five measurements were taken for each homogenate.

Extractions of yamu muscle tissue proteins

The protein extraction was done by modifying the method proposed by Cao *et al.* (2006). A one gram sample was homogenized in a homogenizer (Ultra-turrax IKA T-25, Campinas, Brazil) with 4 volumes of distilled-deionized water for 30 seconds, then centrifuged at 5,000 g and 4°C for 5 min; the supernatant was removed and the precipitate was washed with a cold high ionic strength solution (50 mM phosphate buffer, pH 7.5) for 30 s, then the homogenate was centrifuged at 5,000 g and 4°C for 15 min; the precipitate was suspended in 4 volumes of cold 50 mM phosphate buffer, cold pH 7.5 and repeated the centrifugation process two more times. The supernatant was discarded, a final centrifugation was performed at 3,000 g for 15 min and the precipitate was diluted with 50 mM phosphate buffer and 500 mM NaCl pH 8. The protein extracts were stored at $-40 \pm 0.5^\circ\text{C}$.

Protein quantification

The protein was quantified as suggested by Torres *et al.* (2012), for which 600 mL of each protein extract were used and the required volume of 1% NaCl was added to complete 1 mL; then 1 mL of Bradford reagent was added (Sigma-Aldrich, Saint Louis, MO) to wait for color development for ten minutes and the absorbance was read at 450/590 nm.

SDS-PAGE Electrophoresis (polyacrylamide gel electrophoresis with dodecyl sulfate sodium)

The electrophoresis was done under denaturing conditions (SDS-PAGE) (Mini Vertical Electrophoresis Tetra-PRO-TEAN® Cell, Bio-Rad, Hercules, CA). The separating gel

was 10% and the gel concentration was 5% (w/v). A standard molecular weight of 20 to 210 kDa was used and the staining was done with G-250 colloidal coomassie blue (CBB).

Statistical analysis

The data (pH, texture and WRC) were subjected to an ANOVA to establish whether or not there were differences between the levels of each of the factors. For differences in the levels of the factors, a Tukey test was used. In order to find the effect of the interactions between the factors, an analysis of multifactorial variance was conducted, with the response variables texture (Force cutting), pH and WRC; meanwhile the controlled variables were time and temperature of storage. SPSS 19.0® (2010) was used.

Results

Scanning electron microscopy (SEM)

The scanning electron microscopy analysis of the yamu meat for the effect of cold storage of farmed and fished fillets can be seen in figure 1.

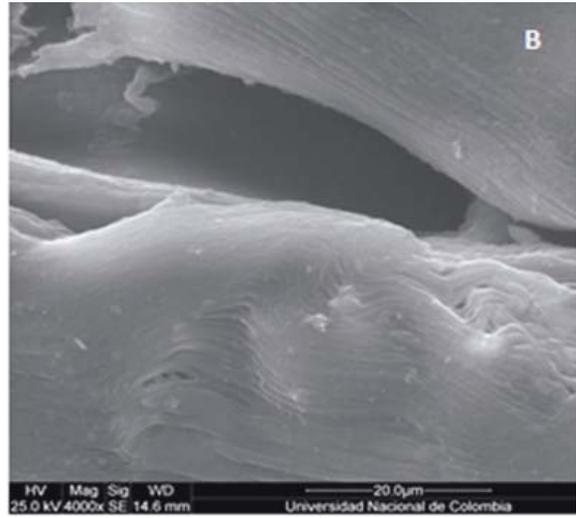
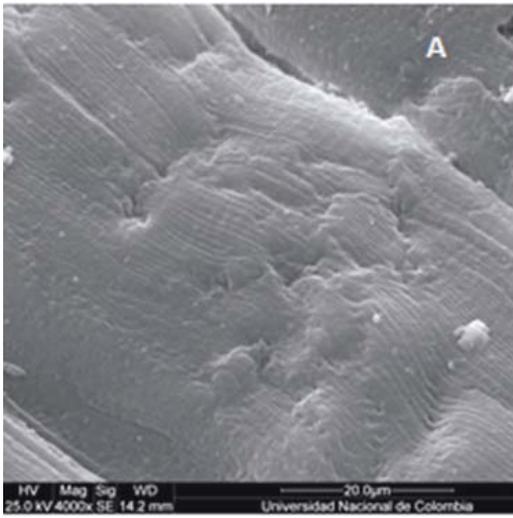
At the microstructural level, the yamu muscle fiber samples that were analyzed when the fish was fresh demonstrated myosepta, myocommata and integral muscle fibers (Fig. 1A and B). Meanwhile, in the samples maintained at 0°C for 13 h (Fig. 3C and D), there were changes in the organization of the muscle fibers, the structure had architectural loss and there was a bond loss between the myotomes. The samples stored at this temperature for 61 h (Fig. 1E and F) showed a greater loss of muscle architecture and collagen degradation. The separation effect between the muscle fibers and general myofibrillar structure presented amorphous forms.

The samples stored at $-8 \pm 1^\circ\text{C}$ (Fig. 1 G, H, I and J) had damage caused by the cold in the muscle structure that was higher; this tissue had pronounced invaginations and the myofibrils lost the original structure because of the effect of the interfiber collagen degradation. This phenomenon is accentuated with increasing the storage period under cold conditions; the samples stored for 61 h had greater detachment between the myosepta and full deformity of the primary structure of the muscle. In addition to this technique, it was possible to see slight differences between the muscular structures of the farmed fish and fish from the river, such as myofibrils and connective tissue. The tissue from the river fish had pronounced fissures starting at 61 hs when stored at $0 \pm 1.5^\circ\text{C}$ (Fig. 1D).

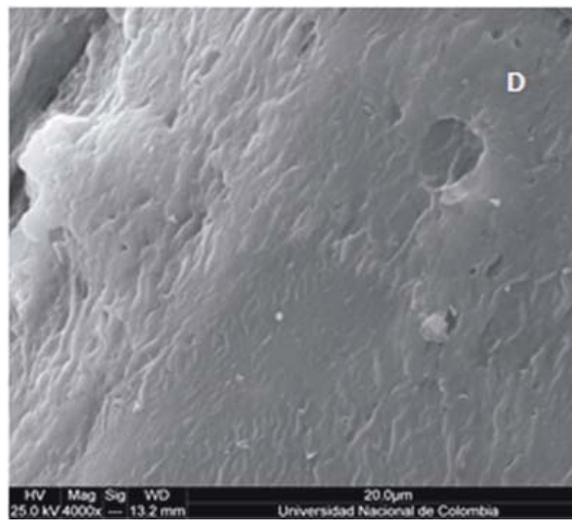
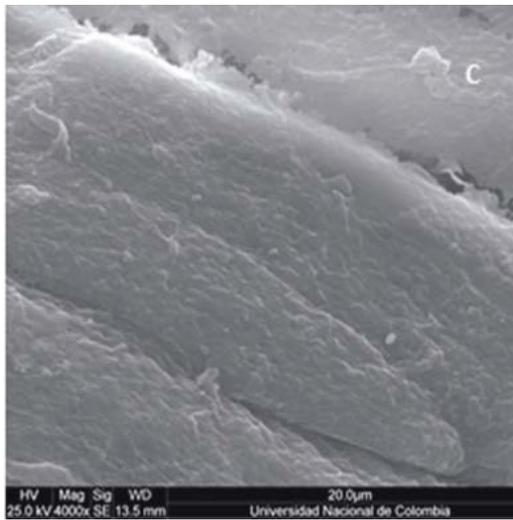
Farmed *Brycon amazonicus*

Fished *Brycon amazonicus*

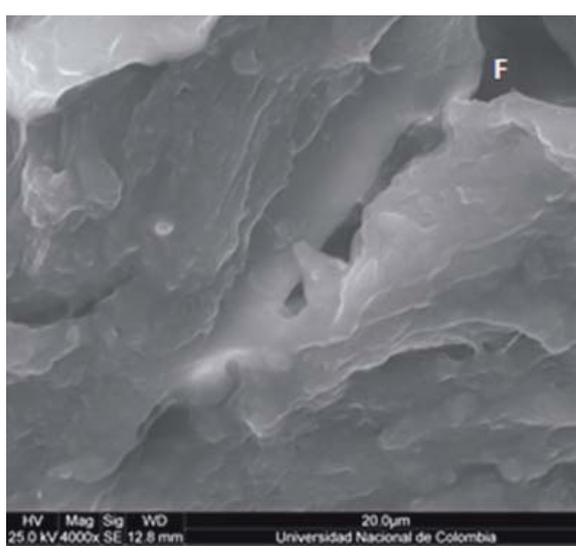
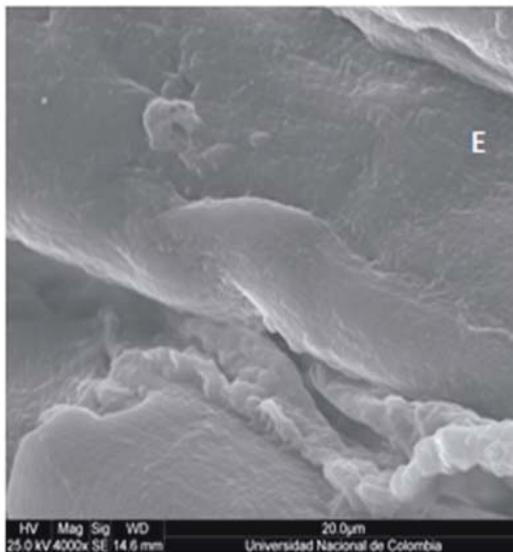
Fresh meat



Meat stored for 3 h at 0±1.5°C



Meat stored for 61 h at 0±1.5°C



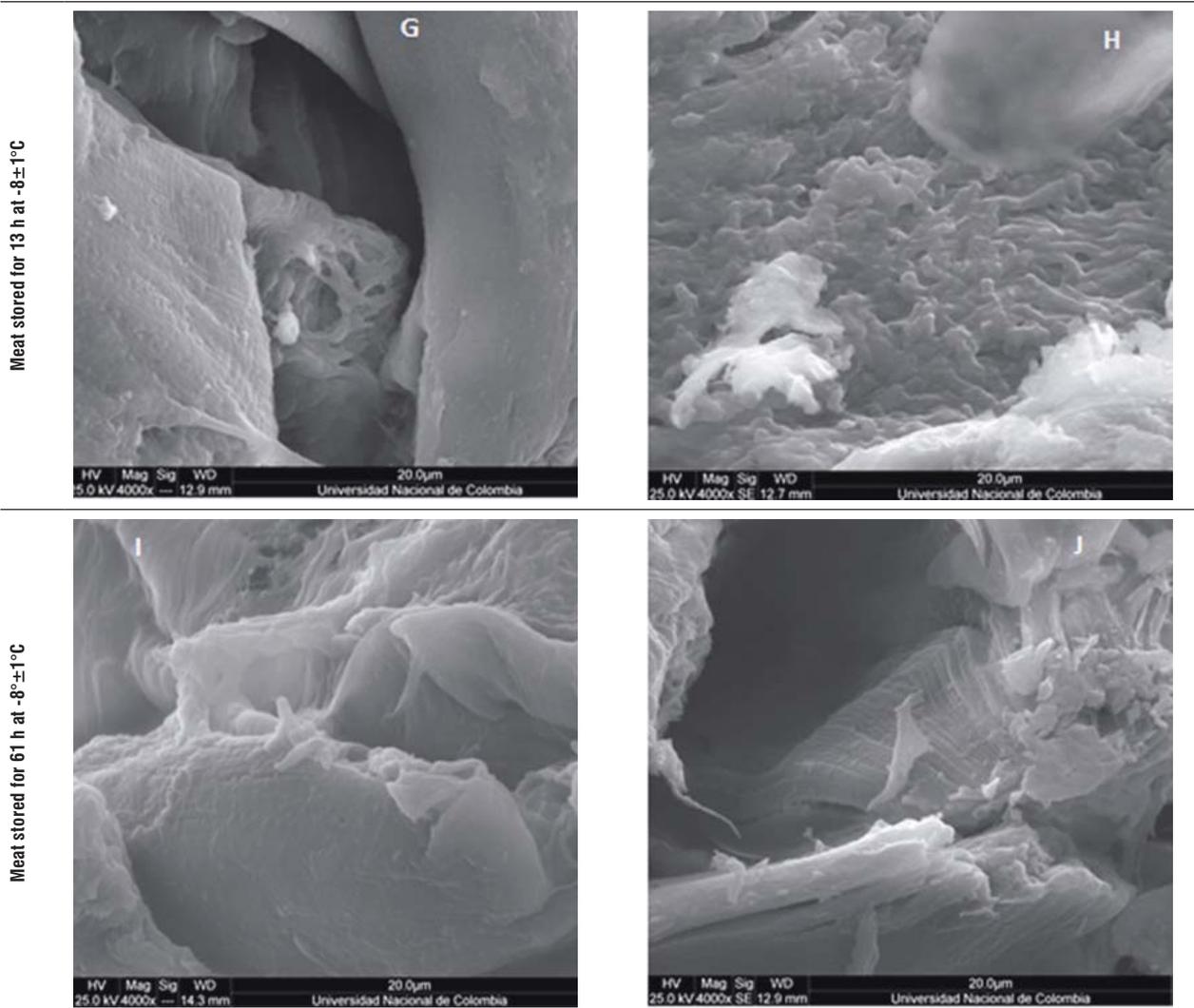


FIGURE 1. Scanning electron microscopy images of *Brycon amazonicus* meat with 4,000 X magnification. A. Fresh farmed meat; B. Fresh fished meat; C. Farmed meat stored for 13 h at $0 \pm 1.5^\circ\text{C}$; D. Fished meat stored for 13 h at $0 \pm 1.5^\circ\text{C}$; E. Farmed meat stored for 61 h at $0 \pm 1.5^\circ\text{C}$; F. Fished meat stored for 61 h at $0 \pm 1.5^\circ\text{C}$; G. Farmed meat stored for 13 h at $-8 \pm 1^\circ\text{C}$; H. Fished meat stored for 13 h at $-8 \pm 1^\circ\text{C}$; I. Farmed meat stored for 61 h at $-8 \pm 1^\circ\text{C}$; J. Fished meat stored for 61 h at $-8 \pm 1^\circ\text{C}$.

Optical microscopy

Figure 2 shows the images of the optical microscopy of the yamu meat (*Brycon amazonicus*) for the effect of cold storage of the fish fillets, from the farm and river.

Fresh meat cuts show a compact structure of muscle fibers, with little separation of endomysium. However; in the myocommata of the river fish, in addition to a mild disorder, an interfibrillar separation effect was observed that was more pronounced in the myocommata than in the farmed fish (Fig. 2A and B). The effect of the low temperature and storage time on the tissue was evidenced by the

degradation of larger areas. When the samples were stored for 13 h at 0°C (Fig. 2C and D), the separation between the endomysium, myosepta and myotomes (arrows) was higher than in the fresh samples, showing a minor effect on the river fish, probably because these were older, factor that affects collagen composition. Moreover, when the cold storage time was equal to 61 h (Fig. 2E and F), the tissue from the farmed fish showed greater involvement in the histological structure and myocommata, showing almost total degradation. While the muscle samples of the river fish, although they had interfibrillar collagen loss, had a defined structure.

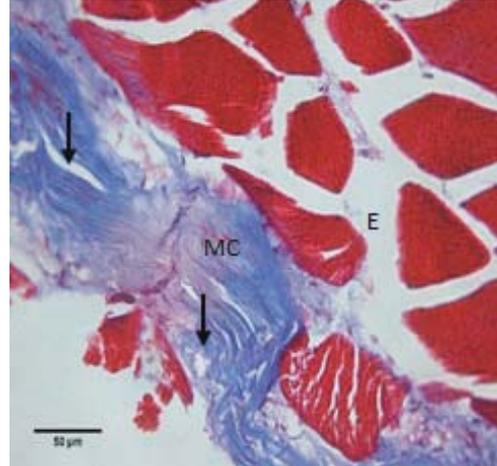
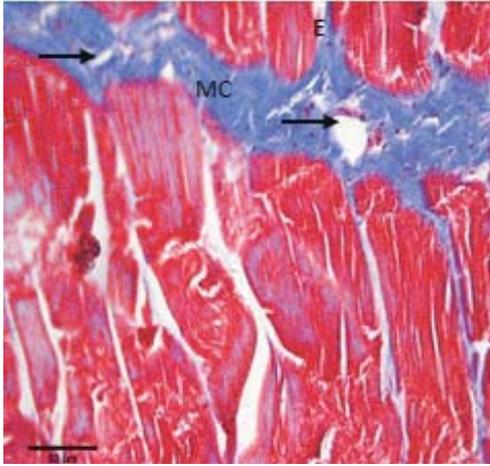
Farmed *Brycon amazonicus*

Fished *Brycon amazonicus*

A

B

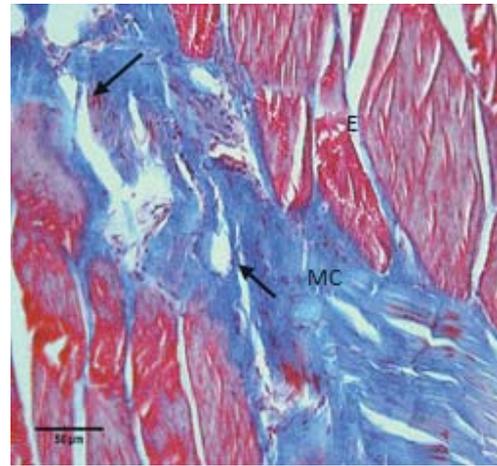
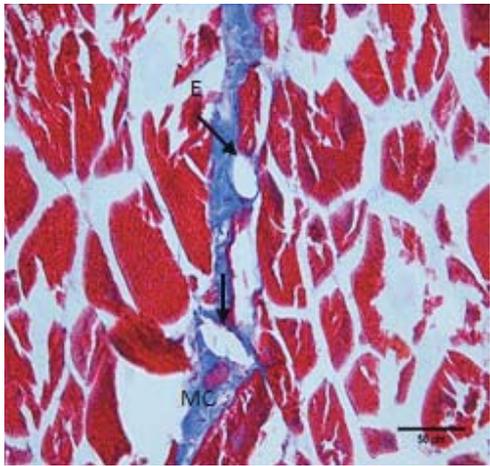
Fresh meat



C

D

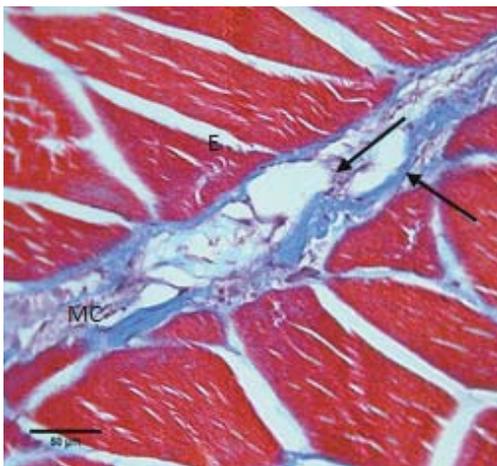
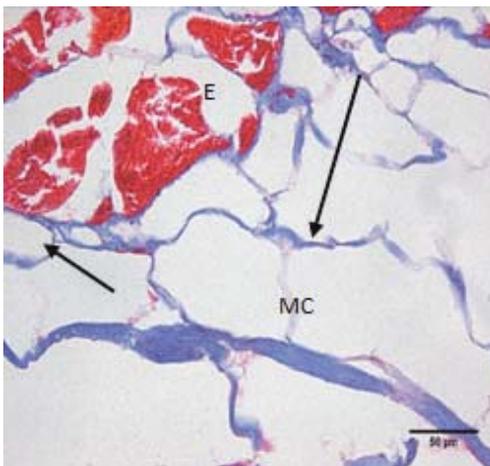
Stored for 13 h at 0±1.5°C



E

F

Stored for 61 h at 0±1.5°C



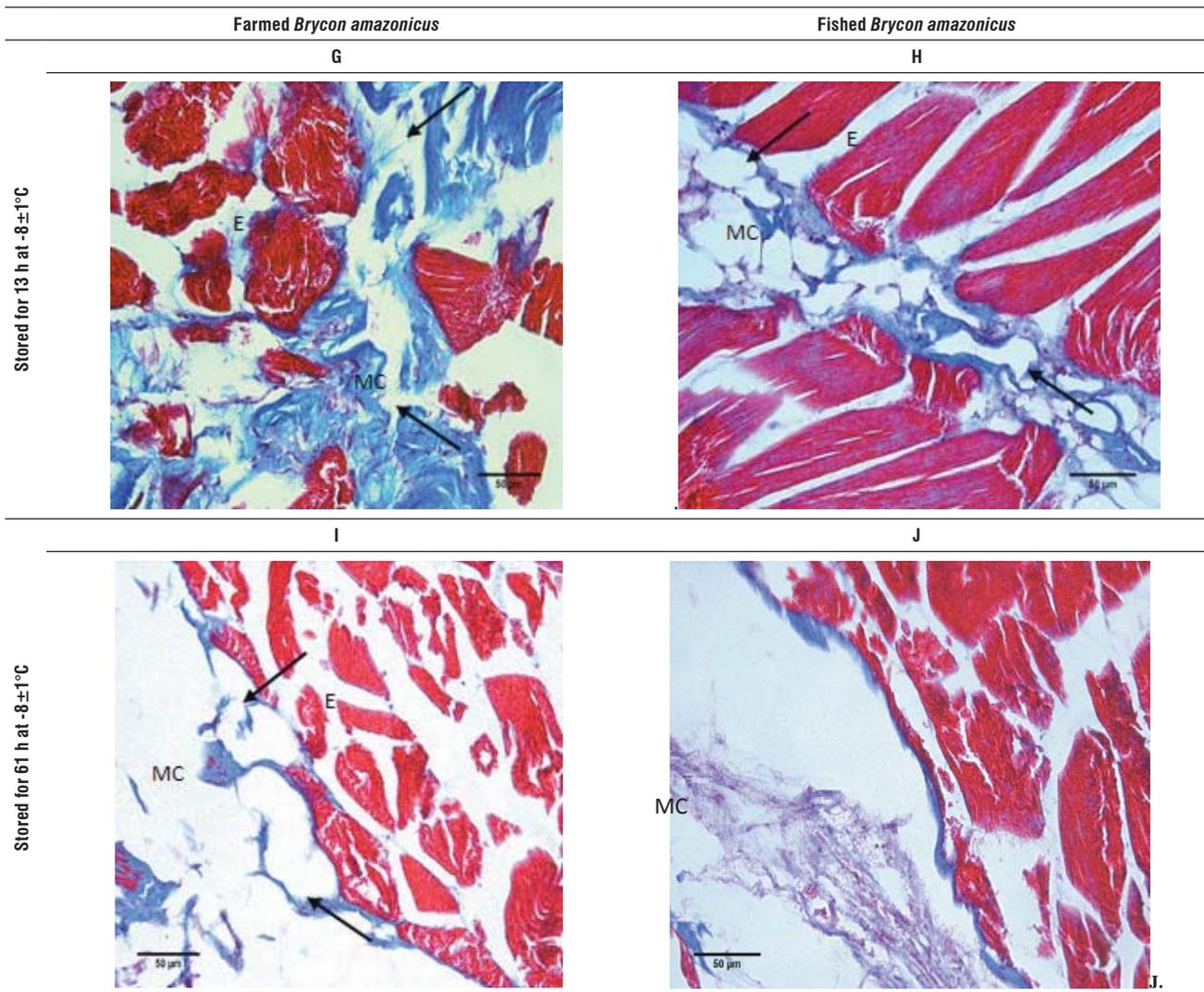


FIGURE 2. Optical microscopy images of yamu *Brycon amazonicus* meat stained with Mason trichrome, 40X magnification. A. Fresh meat from farming; B) Fresh meat from river; C) Farmed meat stored for 13 h at $0\pm 1.5^{\circ}\text{C}$; D. Meat from the river stored for 13 h at $0\pm 1.5^{\circ}\text{C}$; E. Farmed meat stored for 61 h at $0\pm 1.5^{\circ}\text{C}$; F. Meat from the river stored for 61 h at $0\pm 1.5^{\circ}\text{C}$; G. Farmed meat stored for 13 h at $-8\pm 1^{\circ}\text{C}$; H. Meat from the river stored for 13 h at $-8\pm 1^{\circ}\text{C}$; I. Farmed meat stored for 61 h at $-8\pm 1^{\circ}\text{C}$; J. Meat from the river stored for 61 h at -8°C . Ruptured areas are marked with arrows, MC is myocommata and E is endomysium.

Water retention capacity (WRC) of yamu meat samples

The water retention capacity values (WRC) of the yamu meat under different storage conditions can be seen in figure 3.

The WRC of the yamu meat was affected by the storage time ($P\leq 0.05$) where the highest value, besides the value for the fresh meat, was seen in the fillets of the river fish, stored for 13 h at $0\pm 1.5^{\circ}\text{C}$ and the lowest value was seen

in the river fish fillets stored for 61 h at $0\pm 1.5^{\circ}\text{C}$ (29.7%). Moreover, the origin of the fish (river or farmed) and storage temperature treatment had no impact on the variation of the WRC ($P>0.05$); the interactions between the treatments did not have an effect on the WRC either (Fig. 3).

Texture analysis of the yamu meat

The texture results for the yamu meat under different storage conditions are in figure 4.

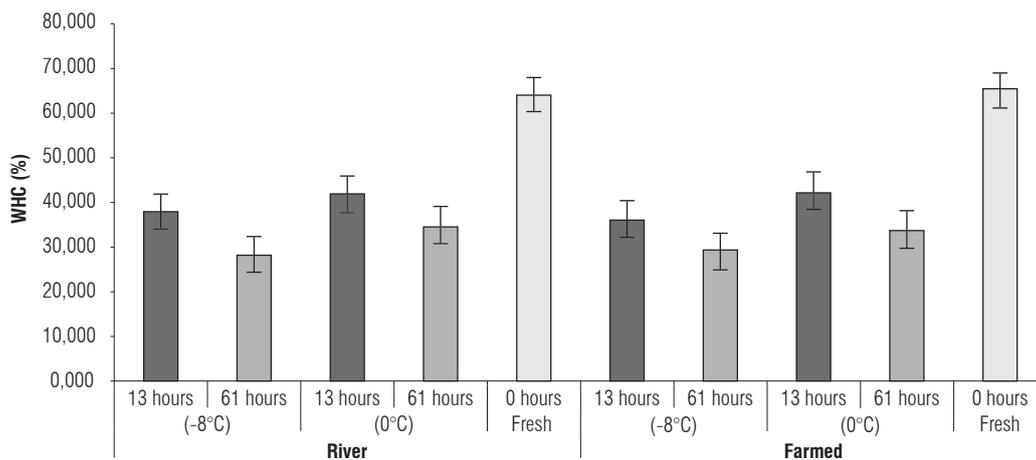


FIGURE 3. Water retention capacity (WRC) of the yamu meat subjected to different storage conditions. The errors bars correspond to standard deviation.

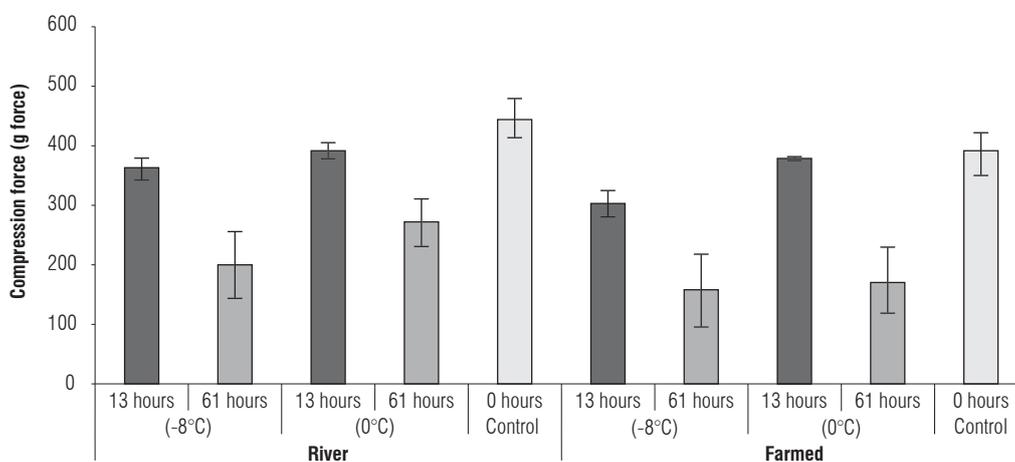


FIGURE 4. Texture analysis of the yamu meat subjected to different storage conditions. The errors bars correspond to standard deviation.

The values indicated that the texture of the yamu meat was affected both by the origin of the fish and by the cold storage period ($P \leq 0.05$); there was also a significant effect from the interaction of the 3 factors on the texture of meat. The origin only affected the texture when the storage time reached 61 h. At 13 h of storage, there were no significant differences ($P > 0.05$) for texture between the farmed fish and the river fish.

Determination of pH in the yamu meat

The analysis of the pH values of yamu fillets stored under different conditions are in figure 5.

The pH values were affected by two of the three factors (origin and treatment time), as well as by the interaction of all factors ($P \leq 0.05$). However, the origin of the fish had an important effect on the pH variation of the fillets after storage for 61 h.

The values showed that the pH of the farmed fish meat ranged from 6.54 ± 0.07 to 6.37 ± 0.03 when stored for 13 and 61 h, respectively. The river fish had a pH of between 6.51 ± 0.08 and 6.48 ± 0.09 for fish preserved at 13 and 61 h, respectively.

Protein degradation

The protein profile analysis of tissue extracts of yamu meat (*Brycon amazonicus*) under different storage conditions is in Fig. 6 and 7.

The electrophoresis showed 14 bands. Slight degradation was observed of these proteins in the samples with cold storage (Fig. 7A), with a greater effect on the samples preserved for 61 h at a lower temperature ($-8 \pm 1^\circ\text{C}$) (lanes 5 and 6). The 222 kDa band (Fig. 7B, lane 2), similar to the heavy chain myosin, was in all of the treatments and, although the intensity of the band decreased, lower molecular weight

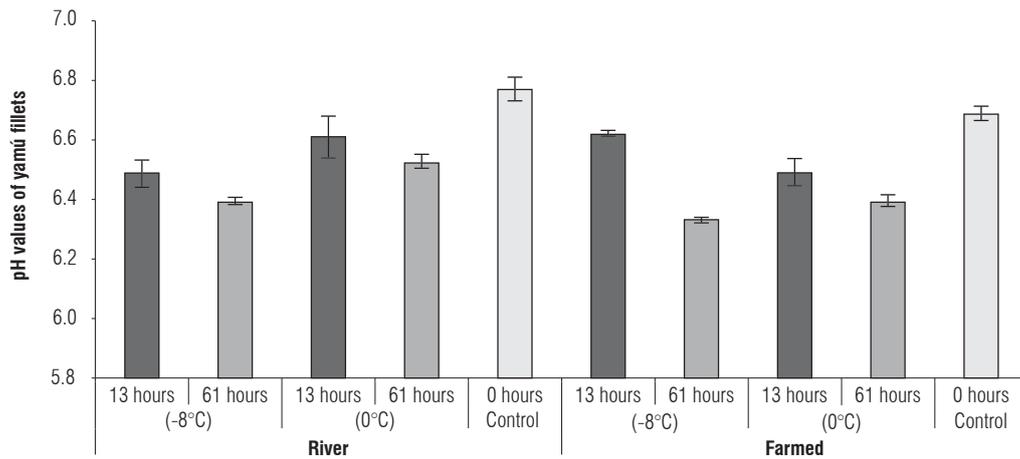


FIGURE 5. pH variation in the yamu fillets stored at different temperatures. The errors bars correspond to standard deviation.

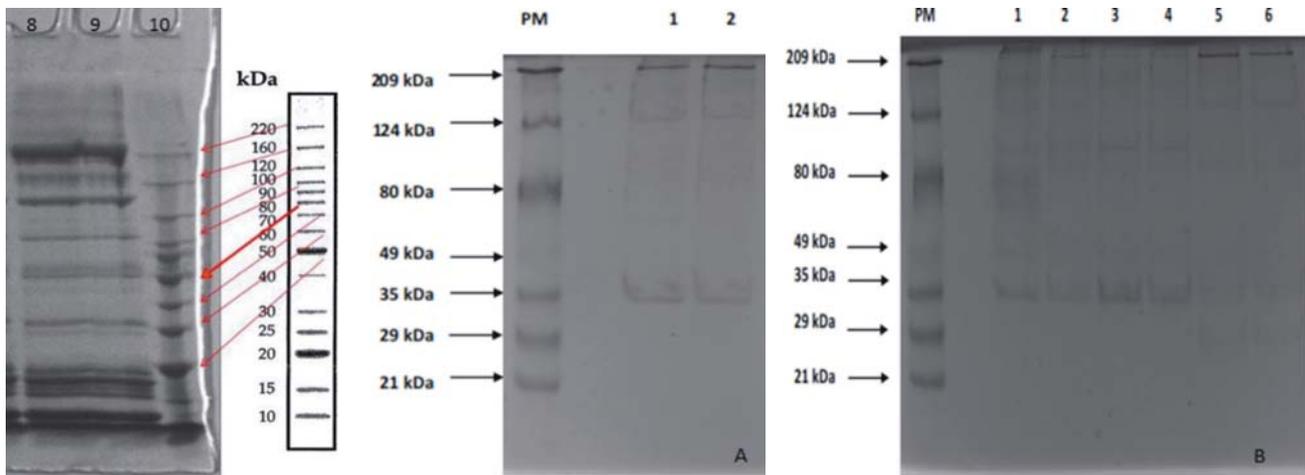


FIGURE 6. Protein profile of tissue extracts of yamu meat (*Brycon amazonicus*). Lane 8: protein profile of *Brycon amazonicus* from the river, lane 9: protein profile of farmed *Brycon amazonicus*.

FIGURE 7. 10% SDS-PAGE Electrophoresis. Colloidal Coomassie was used for staining; 30 μ g of protein were placed in each lane. Fig. 7A shows the protein profiles of the samples stored at $0 \pm 1.5^\circ\text{C}$ for 13 h. PM lane: molecular weight marker. Lane 1: Fresh meat from the river; lane 2: farmed fresh meat. Fig. 7B: PM lane: molecular weight marker; lane 1: meat from the river stored at $0 \pm 1.5^\circ\text{C}$ for 61 h; lane 2: farmed meat stored at $0 \pm 1.5^\circ\text{C}$ for 61 h; lane 3: meat from the river stored at $-8 \pm 1^\circ\text{C}$ for 13 h; lane 4: farmed meat stored at $-8 \pm 1^\circ\text{C}$ for 13 h; lane 5: meat from the river stored at $-8 \pm 1^\circ\text{C}$ for 61 h; lane 6: farmed meat stored at $-8 \pm 1^\circ\text{C}$ for 61 h.

bands were seen (212.69 kDa; 172.87 kDa; 171.96 kDa; 147.05 kDa; 135.96 kDa, 124.32 kDa, and 112.44 kDa) in the samples with cold storage. In this sense, it is possible to conclude that there was no complete degradation. In the samples preserved at $0 \pm 1.5^\circ\text{C}$ for 13 h, there was not a defined band of a molecular weight similar to that of actin (~ 45 kDa), but the appearance of 34.06 kDa and 30.30 kDa bands evidenced a phenomenon suggesting actin degradation. Bands representing α -actinin (~ 105 kDa) stopped appearing after storage for 13 h (Fig. 7A and 7B); however, a similar weight band appeared in the samples of the river fish and farmed fish stored at $0 \pm 1.5^\circ\text{C}$ for 61 h and -8°C for 13 h, respectively. This band did not have the intensity of

the actin bands reported in the literature or found in the initial profiles, indicating that it is a fragment of another protein with a higher molecular weight.

Discussion

The scanning electron microscopy and optical analysis results showed that, at a lower temperature, the effect on the *Brycon amazonicus* muscle tissue was greater. The loss of collagen and the formation of interfiber spaces and their size were the effect of low temperatures, showing significant difference between the treatments when comparing the samples stored at $-8 \pm 1^\circ\text{C}$ and 0°C during the same storage

time. The samples stored at $-8\pm 1^\circ\text{C}$ (Fig. 2 and 3) showed a loss of myocommata and interfibrillar tissue and the formation of spaces in the myocommata and between the myomeres. At this temperature, the effect of the origin of the fish was irrelevant since the chilling effect was dramatic and similar in all of the fish. This phenomenon of degradation of myofibrils connective tissue in *Brycon amazonicus* meat subjected to processes of cold storage is a phenomenon that has been reported previously for different species of fish with cold storage at different temperatures and for different times (Caballero *et al.*, 2009; Ofstad *et al.*, 2006; Papa *et al.*, 1997; Taylor *et al.*, 2002) and is explained as an effect of proteoglycan degradation of the matrix, where the cause is the structural decomposition of the collagen network, generating muscle structure failure (Ofstad *et al.*, 2006). Similar results were reported by others authors for sea bream fillets (*Sparus aurata*) stored at 4°C for 2 d and for salmon fillets (*Salmo salar* L.) kept on ice for 1 d (Taylor *et al.*, 2002).

For the water retention capacity, the values differed from those obtained for European hake fillets (*Merluccius merluccius*) stored at $-40\pm 1.5^\circ\text{C}$ for 25 weeks, where the water retention capacity, although reduced in capacity, remained above 40% (Sánchez-Alonso *et al.*, 2012). There were differences in the values obtained for this species, stored at different temperatures for different periods of time (0-48 weeks), which reported that the effect of storage time also affected the WRC of that species (Sánchez-Valencia *et al.*, 2014). The fact that the WRC values reported by these authors are greater than those obtained in our research clearly shows that the WRC of yamu meat undergoes dramatic changes when it is subjected to cold storage processes, especially at temperatures below $0\pm 1.5^\circ\text{C}$. This phenomenon is caused by alterations in the protein/water caused by exposure of the aliphatic groups in the protein secondary structure complex (Careche *et al.*, 1999). These events occur as a result of protein degradation that is not controlled with storage temperature, which clearly explains why only the storage time causes a reduction in the WRC because, although the fillets were kept at temperatures below 0°C , the enzymatic action that hydrolyzes the protein complex that helps retain water still remains active.

The texture analysis of the yamu meat showed values obtained with the compression force of the yamu meat that were higher than those reported for trout (*Oncorhynchus mykiss*) kept on ice for 5 d and which were between 32 to 116 (g-force) (Godiksen *et al.*, 2009); for *Brycon cephalus*, after storage at -3°C for 12 h, values between 96 and 128 (g-force) were reported (Suárez *et al.*, 2007) and for six

distinct species stored at 5°C for 72 h (Ando *et al.*, 1999). However, these values are lower than those reported for Atlantic salmon (*Salmo salar*) frozen and chilled pre and post mortem rigor, which ranged between 846.4 and 1070.7 (g-force) (Einen *et al.*, 2002). Likewise, the values of this research coincide with those reported by Liu *et al.* (2013), who reported average values of 350 and 430 g for grass carp meat stored at -3°C and 0°C , respectively. Thus, the yamu meat showed a more drastic change than other authors reported, coinciding with Suárez-Mahecha *et al.* (2007), indicating that the meat of animals from the *Brycon* genus can be highly affected by cold storage, more so than the meat of other fish species. This loss of texture is highly related to the decrease in the WRC because hydrolyzed myofibrillar proteins lose the structure that gives support and rigidity to the meat and in effect, the resistance to compression analysis is reduced.

The pH values were different from those reported by other authors (Jain *et al.*, 2007; Ocaño-Higuera *et al.*, 2009) who worked with rohu (*Labeo rohita*) and dogfish shark (*Mustelus lunulatus*) stored on ice for 8 and 18 d, respectively. These researchers showed that the pH of the meat of these species increased as storage time passed; however, in our research, the pH of the meat decreased over time. In addition, the results were similar to that reported for cod (*Gadus morhua*) kept at 0, 5, 10, and 15°C for 400 h (Heising, 2014). This decrease in pH suggests that the fish were subjected to cold storage before rigor mortis began and the decline was the result of the conversion of glycogen to lactate during postmortem glycolysis (Einen *et al.*, 2002).

Biochemical changes play an important role in the texture of fish fillets. In particular, the rapid post-mortem acidification of anaerobic glycolysis and a low final pH have been associated with a loss of texture (Ang and Haard, 1985; Kiessling *et al.*, 2004), possibly because of the reduced strength of the connective tissue, the denaturation of proteins and increased proteolysis (Ofstad *et al.*, 2006).

The behavior of myofibrillar protein analysis of *Brycon amazonicus* with cold storage (0°C and -8°C) was similar for the river fish and the farmed fish, but by storing the fillets longer and at lower temperatures, the appearance of lower molecular weight bands (71.08 and 64.78 kDa) was evident, suggesting some degradation of higher molecular weight proteins, such as troponin (~78 kDa) (Fig. 7b, lanes 3-6).

The protein patterns were similar to those reported by other authors (Ball and Johnston, 1996; Ladrat *et al.*, 2003; Lund and Nielsen, 2001; Ogata *et al.*, 1998; Swartz *et al.*,

1997), within which proteins have been reported, such as heavy chain myosin (WHC ~200 kDa) (Cao *et al.*, 2006; Liu *et al.*, 2013), α -actinin (~105 kDa) (Ladrat *et al.*, 2003), actin (~45 kDa) (Cao *et al.*, 2006) and troponin (~78 kDa) (Ladrat *et al.*, 2003).

Conclusions

There was degradation of proteins, such as heavy chain myosin, actin and α -actinin, in the cold storage of the yamu meat. In addition, the origin of the fish and the storage temperature had a significant effect on the texture changes, and the storage time had a significant effect on the three technological characteristics of the meat. The storage temperature did not significantly affect the water retention capacity.

Literature cited

- Ando, M., A. Nishiyabu, Y. Tsukamasa, and Y. Makinodan. 1999. Post-mortem softening of fish muscle during chilled storage as affected by bleeding. *J. Food Sci.* 64(3), 423-428. Doi:10.1111/j.1365-2621.1999.tb15056.x
- Ang, J. F. and N. F. Haard. 1985. Chemical composition and post-mortem changes in soft textured muscle from intensely feeding atlantic cod (*Gadus morhua*, L.). *J. Food Biochem.* 9(1), 49-64. Doi: 10.1111/j.1745-4514.1985.tb00338.x
- Arias, J. 2006. Estado actual del conocimiento sobre el yamú, *Brycon amazonicus*. *Rev. Colomb. Cienc. Pecu.* 19(2), 125-133.
- Ball, D., and I. Johnston, 1996. Molecular mechanisms underlying the plasticity of muscle contractile properties with temperature acclimation in the marine fish *Myoxocephalus scorpius*. *J. Exp. Biol.* 199(Pt 6), 1363-1373.
- Caballero, M.J., M. Betancor, J.C. Escrig, D. Montero, A. Espinosa de los Monteros, P. Castro, and M. Izquierdo, 2009. Post mortem changes produced in the muscle of sea bream (*Sparus aurata*) during ice storage. *Aquaculture* 291(3-4), 210-216. Doi: 10.1016/j.aquaculture.2009.03.032
- Cao, M.-J., X.-J. Jiang, H.-C. Zhong, Z.-J. Zhang, and W.J. Su, 2006. Degradation of myofibrillar proteins by a myofibril-bound serine proteinase in the skeletal muscle of crucian carp (*Carasius auratus*). *Food Chem.* 94(1), 7-13. Doi: 10.1016/j.foodchem.2004.09.016
- Careche, M., A.M. Herrero, A. Rodriguez-Casado, M.L. Del Mazo, and P. Carmona. 1999. Structural changes of hake (*Merluccius merluccius* L.) fillets: Effects of freezing and frozen storage. *J. Agric. Food Chem.* 47(3), 952-959. Doi: 10.1021/jf9809481
- Chéret, R., C. Delbarre-Ladrat, M.D. Lamballerie-Anton, and V. Verrez-Bagnis. 2007. Calpain and cathepsin activities in post mortem fish and meat muscles. *Food Chem.* 101(4), 1474-1479. Doi: 10.1016/j.foodchem.2006.04.023c
- Einen, O., T. Guerin, O.S. Fjæra, Skjervold, and P.O. 2002. Freezing of pre-rigor fillets of Atlantic salmon. *Aquaculture* 212(1-4), 129-140. Doi: 10.1016/S0044-8486(01)00874-2
- Godiksen, H., M. Morzel, G. Hyldig, and F. Jessen. 2009. Contribution of cathepsins B, L and D to muscle protein profiles correlated with texture in rainbow trout (*Oncorhynchus mykiss*). *Food Chem.* 113(4), 889-896. Doi: 10.1016/j.foodchem.2008.08.012
- Heising, J. K., P. V. Bartels, M. A. van Boekel, and M. Dekker. 2014. Non-destructive sensing of the freshness of packed cod fish using conductivity and pH electrodes. *J. Food Eng.* 124(0), 80-85. Doi: 10.1016/j.jfoodeng.2013.10.008
- Jain, D., P.B. Pathare, and M.R. Manikantan. 2007. Evaluation of texture parameters of rohu fish (*Labeo rohita*) during iced storage. *J. Food Eng.* 81(2), 336-340. Doi: 10.1016/j.jfoodeng.2006.11.006
- Kiessling, A., M. Espe, K. Ruohonen, and T. Mørkøre. 2004. Texture, gaping and colour of fresh and frozen Atlantic salmon flesh as affected by pre-slaughter iso-eugenol or CO₂ anaesthesia. *Aquaculture* 236(1-4), 645-657. Doi: 10.1016/j.aquaculture.2004.02.030
- Ladrat, C., V. Verrez-Bagnis, J. Noël, and J. Fleurence. 2003. *In vitro* proteolysis of myofibrillar and sarcoplasmic proteins of white muscle of sea bass (*Dicentrarchus labrax* L.): Effects of cathepsins B, D and L. *Food Chem.* 81(4), 517-525. Doi: 10.1016/S0308-8146(02)00481-8
- Larsson, T., E.O. Koppang, M. Espe, B.F. Terjesen, A. Krasnov, H., M. Moreno, and T. Mørkøre. 2014. Fillet quality and health of Atlantic salmon (*Salmo salar* L.) fed a diet supplemented with glutamate. *Aquaculture* 426-427, 288-295. Doi: 10.1016/j.aquaculture.2014.01.034
- Liu, D., L. Liang, W. Xia, J.M. Regenstien, and P. Zhou. 2013. Biochemical and physical changes of grass carp (*Ctenopharyngodon idella*) fillets stored at -3 and 0°C. *Food Chem.* 140(1-2), 105-114. Doi: 10.1016/j.foodchem.2013.02.034
- Lund, K.E. and H.H. Nielsen. 2001. Proteolysis in salmon (*Salmo salar*) during cold storage; effects of storage time and smoking process. *J. Food Biochem.* 25(5), 379-395. Doi: 10.1111/j.1745-4514.2001.tb00747.x
- Ma, B. and H. Yamanaka. 1991. Studies on thaw-rigor in red sea bream and carpa muscles. *Nippon Suisan Gakkaishi* 57, 4.
- Mohan, M., D. Ramachandran, T.V. Sankar, and R. Anandan. 2007. Influence of pH on the solubility and conformational characteristics of muscle proteins from mullet (*Mugil cephalus*). *Process Biochem.* 42(7), 1056-1062. Doi: 10.1016/j.procbio.2007.04.005
- Morrison, E.H., H.A. Bremner, and P.P. Purslow. 2000. Location of and post-mortem changes in some cytoskeletal proteins in pork and cod muscle. *J. Sci. Food Agric.* 80(6), 691-697. Doi: 10.1002/(SICI)1097-0010(20000501)80:6<691::AID-JSFA590>3.0.CO;2-T
- Ocaño-Higuera, V.M., E. Marquez-Ríos, M. Canizales-Dávila, F.J. Castillo-Yáñez, R. Pacheco-Aguilar, M.E. Lugo-Sánchez, and A.Z. Graciano-Verdugo. 2009. Postmortem changes in cazon fish muscle stored on ice. *Food Chem.* 116(4), 933-938. Doi: 10.1016/j.foodchem.2009.03.049
- Ofstad, R., R.L. Olsen, R. Taylor, and K.O. Hannesson. 2006. Breakdown of intramuscular connective tissue in cod (*Gadus morhua* L.) and spotted wolffish (*Anarhichas minor* O.) related to gaping. *LWT-Food Sci. Technol.* 39(10), 1143-1154. Doi: 10.1016/j.lwt.2005.06.019

- Ogata, H., F. Aranishi, K. Hara, K. Osatomi, and T. Ishihara. 1998. Proteolytic degradation of myofibrillar components by carp cathepsin L. *J. Sci. Food Agric.* 76(4), 499-504. Doi: 10.1002/(SICI)1097-0010(199804)76:4<499::AID-JSFA980>3.0.CO;2-W
- Oka, H., K. Ohno, and J. Ninomuya, 1990. Changes in texture during cold storage of cultured yellowtail meat prepared by different killing methods. *Bull. Japan Soc. Sci. Fish* 56, 1673-1678. Doi: 10.2331/suisan.56.1673
- Papa, I., R.G. Taylor, C. Astier, F. Ventre, M.C. Lebart, C. Roustan, and Y. Benyamin. 1997. Dystrophin cleavage and sarcolemma detachment are early post mortem changes on bass (*Dicentrarchus labrax*) white muscle. *J. Food Sci.* 62(5), 917-921. Doi: 10.1111/j.1365-2621.1997.tb15006.x
- Simeonidou, S., A. Govaris, and K. Varelziz. 1997. Effect of frozen storage on the quality of whole fish and fillets of horse mackerel (*Trachurus trachurus*) and mediterranean hake (*Merluccius mediterraneus*). *Z. Lebensm. Unters. For. A* 204(6), 405-410. Doi: 10.1007/s002170050102
- Suárez-Mahecha, H., L.H. Beirão, A. De Francisco, L.S.O. Nakaghi, and S.C. Pardo-Carrasco. 2007. Chilling effect on the post-mortem texture of the matrinxã fish muscle *Brycon cephalus*. *Arq. Bras. Med. Vet. Zootec.* 59(4), 1067-1073.
- Suárez-Mahecha, H., A. De Francisco, L.H. Beirao, S. Pardo, and M. Cortés. 2007. Pérdida de textura post mortem de la carne de pescado durante el almacenamiento en frío. *Acta Biol. Colomb.* 12(1), 13-18.
- Swartz, D.R., R.L. Moss, and M.L. Greaser. 1997. Characteristics of troponin C binding to the myofibrillar thin filament: extraction of troponin C is not random along the length of the thin filament. *Biophys J.* 73(1), 293-305. Doi:10.1016/S0006-3495(97)78070-6
- Sánchez-Alonso, I., I. Martínez, J. Sánchez-Valencia, and M. Careche. 2012. Estimation of freezing storage time and quality changes in hake (*Merluccius merluccius* L.) by low field NMR. *Food Chem.* 135(3), 1626-1634. Doi: 10.1016/j.foodchem.2012.06.038
- Sánchez-Valencia, J., I. Sánchez-Alonso, I. Martínez, and M. Careche. 2014. Estimation of frozen storage time or temperature by kinetic modeling of the Kramer shear resistance and water holding capacity (WHC) of hake (*Merluccius merluccius* L.) muscle. *J. Food Eng.* 120(0), 37-43. Doi: 10.1016/j.jfoodeng.2013.07.012
- Taylor, R.G., S.O. Fjaera, and P.O. Skjervold. 2002. Salmon fillet texture is determined by myofiber-myofiber and myofiber-myocommata attachment. *J. Food Sci.* 67(6), 2067-2071. Doi: 10.1111/j.1365-2621.2002.tb09502.x
- Torres L., G.A., I.C. Sánchez B., L.P. Restrepo S., and W. Albarracín H. 2012. Estudio de la maduración de carne de cordero empleando electroforesis sds-page. *Rev. Colomb. Quím.* 41(2), 263-281.
- Toyohara, H. and Y. Shimizu. 1988. Relation of the rigor mortis of fish body and the texture of the muscle. *Nippon Suisan Gakkaishi* 54(10), 1795-1798.
- Useche, C., R. Hurtado, and P. Cala. 1993. Sobre la ecología de *Brycon siebenthalae* y *Milossoma duriventris* (Pices: Characidae) en el río Cafre, Orinoquia. *Rev. Caldasia* 17(2), 341-352.
- Verrez-Bagnis, V., J. Noel, C. Sautereau, J. Fleurence. 1999. Desmin degradation in postmortem fish muscle. *J. Food Sci.* 64(2), 240-242. Doi: 10.1111/j.1365-2621.1999.tb15873.x
- Wang, P.A., I. Martínez, and R.L. Olsen. 2009. Myosin heavy chain degradation during post mortem storage of Atlantic cod (*Gadus morhua* L.). *Food Chem.* 115(4), 1228-1233. Doi: 10.1016/j.foodchem.2009.01.031
- Wang, P.A., B. Vang, A.M. Pedersen, I. Martínez, and R.L. Olsen. 2011. Post-mortem degradation of myosin heavy chain in intact fish muscle: Effects of pH and enzyme inhibitors. *Food Chem.* 124(3), 1090-1095. Doi: 10.1016/j.foodchem.2010.07.093